Role of vitamin A metabolism in IIH: Results from the idiopathic intracranial hypertension treatment trial

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A B S T R A C T
Introduction: Vitamin A and its metabolites (called retinoids) have been thought to play a role in the development of idiopathic intracranial hypertension (IIH). The IIH Treatment Trial (IIHTT) showed the efficacy of acetazolamide (ACZ) in improving visual field function, papilledema grade, quality of life and cerebrospinal fluid (CSF) pressure. We postulated that IIH patients would demonstrate elevated measures of vitamin A metabolites in the serum and CSF.

Methods: Comprehensive measures of serum vitamin A and its metabolites were obtained from 96 IIHTT subjects, randomly assigned to treatment with ACZ or placebo, and 25 controls with similar gender, age and body mass index (BMI). These included retinol, retinol binding protein, all-trans retinoic acid (ATRA), alpha- and beta-carotenes, and beta-cryptoxanthin. The IIHTT subjects also had CSF and serum vitamin A and metabolite measurements obtained at study entry and at six months.

Results: At study entry, of the vitamin A metabolites only serum ATRA was significantly different in IIHTT subjects (median 4.33 nM) and controls (median 5.04 nM, p = 0.02). The BMI of IIHTT subjects showed mild significant negative correlations with serum ATRA, alpha- and beta-carotene, and beta-cryptoxanthin. In contrast, the control subject BMI correlated only with serum ATRA. At six months, the serum retinol, alpha-carotene, beta-carotene, and CSF retinol were increased from baseline in the ACZ treated group, but only increases in alpha-carotene (p = 0.02) and CSF ATRA (p = 0.04) were significantly greater in the ACZ group compared with the placebo group. No other vitamin A measures were significantly altered over the six months in either treatment group. Weight loss correlated with only with the change in serum beta-carotene (r = −0.44, p = 0.006) and the change in CSF retinol (r = −0.61, p = 0.02).

Conclusion: Vitamin A toxicity is unlikely a contributory factor in the causation of IIH. Our findings differ from those of prior reports in part because of our use of more accurate quantitative methods and measuring vitamin A metabolites in both serum and CSF. ACZ may alter retinoid metabolism in IIH patients.

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1. Introduction

Idiopathic intracranial hypertension (IIH) is a disorder characterized by elevated intracranial pressure with clinical presentation of headaches, pulsatile tinnitus, transient visual obscurations, papilledema and visual loss. The underlying mechanism(s) causing the intracranial hypertension remains unclear with numerous unproven causation theories [1–5]. Obesity is a demonstrated strong risk factor for adult IIH, especially for women of child-bearing age [6–9]. IIH affects children and adults, with a very strong predilection for women after puberty.

Secondary intracranial hypertension with symptoms and findings similar to IIH can be induced by consumption of large quantities or chronic lower doses of retinoid (vitamin A and its natural and synthetic analogs, referred to as retinoids) [8,10–12]. For example, oral ingestion of preformed vitamin A, present in liver and vitamin A related pharmacological therapies have been reported to elevate CSF pressure in adults [13,14], and cause acute protrusion of the fontanel in infants [15]. This has led to the speculation that IIH may be due to the “toxicity of free

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retinoids” formed due to abnormal vitamin A metabolism in obesity. However, continuous intake of vitamin A or its metabolite is required to have unremitting intracranial hypertension and, unlike in IIH, CSF pressure readily normalizes after stopping the inciting agent [12,13].

Vitamin A metabolism is a complicated process and involves multiple enzymes, binding proteins, and receptors (Fig. 1).

Retinol, a major form of vitamin A present in humans and other mammals, is derived from either consumption of food containing preformed vitamin A or via bioconversion from provitamin A carotenoids, including beta-carotene, alpha-carotene, and beta-cryptoxanthin, in the diet [16–18]. Due to its hydrophobic nature, retinol in the circulation is bound to its carrier protein, retinol-binding-protein (RBP), and together with transthyretin (TTR) forms a complex, with the molar ratio of 1:1:1 [13,19]. Retinol is also found in the CSF bound to the TTR/RBP complex. TTR and RBP are produced abundantly in the choroid plexus, the major site of CSF production and macro-molecule delivery to brain. Retinol uptake into target cells is achieved either by free diffusion [20,21] or by a receptor-mediated process through a membrane-bound RBP receptor: STRA6 (stimulated-by-retinoic-acid-6) [22,23]. After uptake into the cell, retinol is metabolized into retinaldehyde and then to all-trans-retinoic acid (ATRA). The meninges and choroid plexus are thought to be the primary sites of ATRA production in the adult brain [24]. ATRA then plays a critical role in regulating gene expression of >500 genes upon its binding to specific nuclear receptors [25–27].

One hypothesis [28] of how retinoid toxicity may lead to IIH is that ATRA increases gene expression of a molecule in the arachnoid granulation cap cells, in ependymal or glial cells, or even in lymphatic pathways, and causes increased resistance to CSF absorption [29,30]. Understanding whether abnormalities in retinoid metabolism may occur in IIH requires measuring retinol levels as well as the transcriptionally active ATRA and its precursors and other retinoid metabolites which can act as antagonists to ATRA. For example, provitamin A carotenoids can be converted to vitamin A through central cleavage by beta-carotene 15,15′-monoxygenase 1 (BCMO1) [31] or can be eccentrically cleaved into beta-apo-carotenoids under the control of beta-carotene-9′,10′-oxygenase (BCO2) [31,32]. Beta-apo-carotenoids can functionally antagonize retinoic acid receptors [33,34]. Therefore, one metabolite of vitamin A can stimulate the retinoic acid receptor and another can antagonize it.

Relationships between vitamin A metabolites, carotenoids and IIH must also be considered in relation to human body mass index (BMI). Vitamin A and its metabolites have been implicated in adipose tissue metabolic pathways and in obesity. In addition, adipose tissue is actively involved in maintaining retinoid homeostasis [35–38]. BMI, as a reflection of fat deposition, correlates positively with the risk of IIH [79] and with the plasma levels of apo-RBP (RBP unbound to retinol), but negatively with plasma carotenoids [39] and serum retinol [40,41].

In this study, we investigated whether plasma vitamin A metabolite and carotenoid levels are elevated in IIH patients enrolled in the Idiopathic Intracranial Hypertension Treatment Trial (IIHTT) compared with obese controls. We also examined the CSF levels of retinoids and of carotenoids in IIH and changes in the serum and CSF over a six month interval when study subjects were treated with acetazolamide (ACZ) or matching placebo. This is the first study to measure serum and CSF ATRA and the first to offer a comprehensive examination of retinoids and provitamin A carotenoids in both serum and CSF. Additionally, since weight management was one of the therapeutic interventions provided to all study subjects, the IIHTT affords us an opportunity to further understand the complex relationships among vitamin A metabolites, carotenoids, and RBP and obesity and weight loss.

2. Methods

2.1. Subjects

Details of the IIHTT study design and entry criteria and outcome are published [42,43]. Newly diagnosed IIH patients naïve to treatment with a perimetric mean deviation (PMD) of −2.00 dB to −7.00 dB using the SITA standard 24-2 test pattern on the Humphrey Field Analyzer II perimeter in the worse eye (‘study eye’) were enrolled. All subjects signed informed consent and the study was performed under institutional review board approval and in accordance with the Helsinki Declaration. Participants were randomized to acetazolamide or placebo for six months, with both treatment groups provided a weight
management and counseling program. The diet program encouraged limited sodium and calories but did not specifically reduce retinoid or carotenoid intake [42]. No study subjects were known to regularly consume a diet high in vitamin A or carotenoids or take vitamin A or carotenoid supplements. Standardized fundus photographs, Frisén grading of photos at the photographic reading center and by clinical examination by site investigators, and high contrast visual acuity and threshold 24-2 perimetry were performed at each visit during the treatment interval of six months. Of the 165 subjects enrolled, 96 participated in this vitamin A study with 25 control subjects without IIH, who had similar age, gender and BMI. Cost considerations prevented the vitamin A study from including all study subjects and a larger number of control subjects as originally planned.

2.2. Laboratory procedures

Retinol, carotenoids (beta-carotene, alpha-carotene, and beta-cryptoxanthin), RBP, and ATRA were measured at baseline (in all subjects) and six months (in IIH subjects). The levels were determined at the Columbia University laboratories using established protocols.

2.2.1. Radioimmunoassay (RIA) for RBP levels

The RIA uses purified human plasma proteins of RBP for iodination by a lactoperoxidase procedure and for standards. A standard displacement curve was established by plotting the percentage of maximal binding of [125]I-RBP with known amounts of purified human plasma RBP for a standard dilution of a rabbit polyclonal antihuman RBP antibody. This RIA procedure has been used previously for measures of RBP in human sera and CSF [41,44–46] and has been previously described in detail [47].

2.2.2. High performance liquid chromatography (HPLC) measurements of retinol and carotenoids

All-trans-retinol and carotenoids (beta-carotene, alpha-carotene, beta-cryptoxanthin, and lycopene) were extracted from 2 ml of CSF or 1 ml of serum after first denaturing proteins through addition of an equal volume of absolute ethanol containing a known amount of retinyl acetate internal standard. Subsequently, 5 ml of hexane, containing a known amount of echinene, an internal standard used for carotenoid determination, was added. Following phase separation by centrifugation, the retinoid and carotenoid containing hexane phase was removed, evaporated under nitrogen, and re-dissolved in benzene for HPLC analysis. The extracted vitamin A metabolites and carotenoids were separated on a 5-μm 4.6 × 250-mm Ultrasphere C18 column preceded by a C18 guard column, using 70% acetonitrile, 15% ethanol, 15% methylene chloride as the running solvent flowing at 1.8 ml/min. Retinol and retinyl esters were detected at 325 nm using a Waters 996 Photodiode Array ultraviolet absorbance monitor. Carotenoids were detected simultaneously at 450 nm. Low limits of detection for retinol, retinyl esters, alpha-carotene, beta-carotene, and beta-cryptoxanthin were respectively, 2, 4, 10, 12, and 6 for plasma-CSF. The assay variability for assays performed on the same day was between 3 and 6%; and between 5 and 8% for assays performed on different days [48,49].

2.2.3. Liquid chromatography tandem mass spectrometry (LC/MS/MS) measurements of ATRA

Owing to the biological and chemical instability of ATRA, plasma and CSF samples were stored in light protected (amber) glass tubes at −80 °C prior to analysis. All LC/MS/MS analyses were carried out within 10 days of sample collection and the great preponderance of these was undertaken within seven days after sample collection. Serum and CSF levels of ATRA were determined by ultra-high performance liquid chromatography tandem mass spectrometry (LC/MS/MS) using a Waters Xevo TQ MS ACQUITY UPLC system (Waters, Milford MA). For this analysis, we only employed LC/MS grade acetonitrile and LC/MS grade water purchased from Thermo Fisher (Pittsburgh, PA).

All-trans-retinoic acid was purchased from Sigma-Aldrich. Penta-deuterated all-trans-retinoic acid (atRA-d5) was employed as an internal standard and was purchased from Toronto Research Chemicals (North York, Ontario). Retinoid concentrations were verified spectrophotometrically using published ε values [40]. Serum and CSF were extracted using the two-step acid-base extraction described by Guenegou et al. [41]. All-trans-retinoic acid was detected and quantified using the multiple reaction monitoring mode (MRM) employing the following transitions: all-trans-retinoic acid, m/z 301.16 → 123.00; and atRA-d5, m/z 306.15 → 127.03. The within-day and between-day coefficients of variation for ATRA analysis were respectively 4.9% and 5.8%.

2.3. Statistical analysis

Wilcoxon rank sum tests were used to compare serum and CSF vitamin A and metabolite values between IIH subjects and controls, as well as to compare 6-month changes in these values (as well as changes in weight) between the ACZ and placebo groups among IIH subjects. Associations between vitamin A and metabolite values and clinical measures (BMI, weight, perimeter mean deviation (PMD)) and Frisén grade of the study (worse) eye, and CSF pressure were summarized using Spearman rank correlations; associations between 6-month changes in these quantities were similarly analyzed. Box plot figures show median (horizontal line), T-bars that extend from the boxes (whiskers) 1.5 times the height of the box, and outlier points.

3. Results

IIH and control subjects had similar distributions of age and BMI (Table 1).

RBP and retinol were also in an approximately 1:1 relationship for both controls and IIH subjects (data not shown), which served as an internal control, demonstrating the expected relative values at baseline and validating the measurements. The serum ATRA level was significantly lower in the IIHTT subjects than in obese controls at baseline, but there were no other group differences regarding baseline values of retinoids or carotenoids (pro-vitamin A and non-pro-vitamin A) at study entry (Table 2).

The BMI of IIHTT subjects showed negative mild to moderate correlations with serum ATRA (r = −0.25, p = 0.01), alpha-carotene (r = −0.35, p = 0.0005), beta-carotene (r = −0.39, p < 0.0001), and beta-cryptoxanthin (r = −0.47, p < 0.0001), but not with serum retinol (r = −0.14, p = 0.17) or RBP (r = 0.05, p = 0.61). In control subjects only the serum ATRA negatively correlated with BMI (r = −0.42, p = 0.04). The clinical features of IIH, papilledema grade, mean deviation of the study eye, and CSF pressure, were not associated with any serum retinoid measurements. The CSF pressure was mildly negatively correlated only with the CSF ATRA (r = −0.27, p = 0.008).

At six months, some vitamin A metabolites increased over baseline in the ACZ treated subjects but not in the placebo treated subjects (Figs. 2–6); these included serum retinol, alpha-carotene, and beta-carotene, and CSF retinol (Table 3).

The BMI and ACZ level were greater than those in the placebo group only for alpha-carotene (p = 0.01, Fig. 3) and CSF ATRA (p = 0.04, Fig. 6). In agreement with previously reported findings [43], ACZ-treated subjects showed greater weight loss (mean − 8.6 ±
7.3 kg) than placebo-treated subjects (mean $-4.5 \pm 6.0$ kg, $p = 0.007$) in this subset of the study cohort. Among all IIHTT subjects, of the serum and CSF retinoid measures, only the change in beta-carotene ($r = -0.44$, $p = 0.006$) and the change in CSF retinol ($r = -0.61$, $p = 0.02$) were correlated with the change in weight. There were no associations between changes in retinoid and provitamin A carotenoid levels in serum or CSF and changes in CSF pressure, Frisén grade or mean deviation of the study eye.

4. Discussion

The data in this study raise doubts about a theory of “toxicity of free retinoids” as a principal pathophysiologic mechanism for IIH. We investigated the baseline and the six month response of retinoids and provitamin A carotenoids in serum and CSF in IIH subjects in a prospective randomized clinical trial and unexpectedly found low serum levels of the transcriptionally active ATRA in IIH subjects at baseline and evidence of a treatment effect of ACZ on retinoid metabolism (in CSF) in IIH subjects. We had hypothesized that we would see elevated levels of vitamin A metabolites either in serum or CSF, with IIH mimicking the intracranial hypertension seen in association with vitamin A toxicity, but we observed no increased measures prior to treatment. Further, there were no differences between IIH subjects and obese controls regarding the intake of the non-provitamin A carotenoids that are abundant in regular diets. The ratio of retinol:RBP in blood, proposed as a measure of the availability of free retinol [50], did not differ significantly at baseline in our study between the IIH patients and the controls. Additionally, the data are only suggestive that ACZ treatment has an impact on vitamin A metabolism in IIH. We could not address whether a weight management program had any effect on vitamin A metabolism since we had no control group without weight management. However, given the known association of exogenous retinoids with secondary intracranial hypertension, reporting the findings of our prospective study and putting them into context of the current knowledge of human vitamin A metabolism is warranted to apply to future investigations.

Our review of the literature found little agreement among previous studies examining retinol levels or RBP levels in serum or CSF in IIH.

Table 2

<table>
<thead>
<tr>
<th></th>
<th>IIH subjects (n = 96)</th>
<th>Control subjects (n = 25)</th>
<th>p-Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serum retinol (μM)</td>
<td>1.56 (1.38, 1.90)</td>
<td>1.58 (1.32, 1.82)</td>
<td>0.71</td>
</tr>
<tr>
<td>Serum ATRA (nM)</td>
<td>4.33 (3.59, 5.18)</td>
<td>5.04 (4.04, 5.84)</td>
<td>0.02</td>
</tr>
<tr>
<td>Serum beta-carotene (μM)</td>
<td>0.21 (0.14, 0.34)</td>
<td>0.21 (0.14, 0.34)</td>
<td>0.96</td>
</tr>
<tr>
<td>Serum alpha-carotene (μM)</td>
<td>0.05 (0.02, 0.07)</td>
<td>0.04 (0.03, 0.09)</td>
<td>0.97</td>
</tr>
<tr>
<td>Serum beta-cryptoxanthin (μM)</td>
<td>0.08 (0.05, 0.12)</td>
<td>0.08 (0.05, 0.10)</td>
<td>0.59</td>
</tr>
<tr>
<td>Serum RBP (μM)</td>
<td>1.59 (1.32, 2.11)</td>
<td>1.62 (1.36, 1.89)</td>
<td>0.98</td>
</tr>
<tr>
<td>Molar retinol:RBP ratio</td>
<td>1.00 (0.85, 1.14)</td>
<td>0.95 (0.89, 1.12)</td>
<td>0.73</td>
</tr>
<tr>
<td>CSF ATRA (pg/ml)</td>
<td>3.17 (2.00, 7.45)</td>
<td>NA</td>
<td></td>
</tr>
<tr>
<td>CSF RBP (nM)</td>
<td>3.40 (2.05, 5.21)</td>
<td>NA</td>
<td></td>
</tr>
<tr>
<td>CSF retinol (nM)</td>
<td>2.37 (2.02, 3.14)</td>
<td>NA</td>
<td></td>
</tr>
</tbody>
</table>

ATRA = all-trans retinoic acid; RBP = retinol-binding-protein; NA = not applicable.

Values are median (25th percentile, 75th percentile); p-values are based on Wilcoxon rank sum tests.

Note, when the p-values are compared to the adjusted significance for multiple comparisons, significance requires $p < 0.007$. 

Fig. 2. Boxplot of serum retinol by treatment group, acetazolamide (ACZ) and placebo, at baseline (Mo. 0) and at six months (Mo. 6). A slight increase in serum retinol level is seen in both groups at six months.

Fig. 3. Boxplot of serum alpha-carotene by treatment group, acetazolamide (ACZ) and placebo, at baseline (Mo. 0) and at six months (Mo. 6). A significant difference is seen between the ACZ and placebo groups with respect to the change in serum alpha-carotenes at six months.

Fig. 4. Boxplot of serum beta-carotene by treatment group, acetazolamide (ACZ) and placebo, at baseline (Mo. 0) and at six months (Mo. 6). An increase in beta-carotene level was seen in both groups, which was greater in the ACZ-treated group, at six months.
patients. Some previous clinical studies showed that the level of serum retinol is elevated in IIH patients [50,51] while other studies found no difference in serum retinol in those with and without IIH [52-54]. One study reported an elevated level of CSF retinol in an individual with IIH [54]. Only one prior study [55], reported as an abstract, measured retinoic acid (total RA and 13-cis-retinoic acid) in the CSF but did not report ATRA in the serum. In a prior study of RBP in IIH, serum RBP was elevated in seven of the 30 IIH patients, but not in any of the 17 control subjects [53].

The variable results across studies may be due to differing methodology with some approaches being less quantitative than the HPLC and LC/MS/MS used in our study. The volume of CSF and serum used for extraction and whether or not the control samples were protected from light also likely contributed to the differences between studies.

Furthermore, patients and controls did not have comparable BMI in all studies. For example, the IIH patients had significantly greater BMI than controls in the Jacobson report [51]. The dietary intake of vitamin A and of provitamin A carotenoids was not accounted for in most studies and there may have been group differences and differences between studies. Our study had a larger enrollment and measured carotenoids and retinoids individually, which may also explain some of the differences in our findings compared with prior reports. Carotenoids levels were the same in IIH patients and control subjects at study entry, suggesting that vitamin A dietary intake was the same between groups.

We also compared retinoids and provitamin A carotenoids in serum and CSF in IIH patients treated with ACZ and compared to study entry and to an IIH patient placebo group. Although serum retinol, beta-carotene, alpha-carotene and CSF retinol were all increased at six months after treatment with ACZ compared with values at study entry, the difference between the ACZ and placebo groups was significant only for the 6-month change in alpha-carotene. Both the ACZ group and the placebo group were placed on weight loss diets and the ACZ group experienced greater weight loss [43]. There is evidence in the literature for a negative correlation between serum carotenoids and human body weight [39]. We cannot determine whether ACZ has an independent impact on carotenoid metabolism given that among all of the IHTT subjects, the increase in serum beta-carotene modestly correlated ($r = -0.44, p = 0.006$) with the weight reduction. In the CSF, there was an unexpected increase in CSF ATRA and retinol levels in the ACZ group at six months, not seen in the placebo group, although the group differences were not statistically significant. The number of subjects in each treatment group was too small to determine whether any effects of ACZ on retinoid levels in serum or CSF were independent of its effect on weight loss. 6-month changes in CSF pressure and papilledema did not appear to be associated with corresponding changes in vitamin A metabolism.

We found the baseline levels of serum ATRA in IIH subjects were low compared to the levels in obese controls. Others have suggested (without supportive data) that serum ATRA would be elevated in IIH patients. This was hypothesized based on the observation of elevated intracranial pressure with vitamin A-related pharmacological therapies such as ATRA used in the treatment of acute promyelocytic leukemia, as well as the previous reports describing elevated serum retinol in IIH in some [50,51] reports. The low serum ATRA in IIH patients, but not in obese control subjects, is suggestive of a perturbation in the retinoid metabolic pathway. Our study is the first study to measure ATRA, which outside of the retina is the major biologically active vitamin A metabolite (see [13] for review). It is of note that, while ACZ treatment plus diet was associated with an increase in the ATRA, there was no similar change with placebo plus diet.

Of all the channels and receptors found in choroid plexus epithelia, aquaporin-1 has raised particular interest in IIH research [56]. Notably, the expression of aquaporin-1 in vitro is increased by ATRA [57] and specific deletion of aquaporin-1 in mice reduces CSF production and intracranial pressure [58]. It is possible that if vitamin A metabolites play a role in IIH pathophysiology or treatment, it is through the transcriptional regulatory actions of ATRA and production of CSF in the choroid plexus [13]. However, there is evidence that retinoic acid may also induce expression of aquaporin-4 in vitro and aquaporin-4 null mice have raised intracranial pressure and ventricular dilation [59]. Aquaporin-4 may be found in ependyma and astrocytes, and there is evidence that these cells take up CSF, possibly balancing the role of aquaporin-1 in the production of CSF in the choroid plexus. Yet another plausible explanation for the effectiveness of ACZ lowering CSF pressure might be an interaction between vitamin A metabolites and aquaporin control of fluid movements [60]. ACZ decreases expression of the AQP1 protein that ordinarily increases CSF production [61]. ACZ also inhibits the hydration of acetaldehyde [62], which is an inhibitor of ATRA production from all-trans-retinol [63]. Given our finding that ACZ raises CSF ATRA levels, the exact mechanism for ACZ effectiveness remains unanswered.
Our study had limitations. Although the distributions of gender and BMI were comparable in IIH subjects and controls, there were fewer controls than IIH subjects. The small size of the control sample in particular resulted in limited power to detect potentially meaningful differences between the IIH and control groups. The controls had no CSF studies and not all IIH subjects had follow-up serum or CSF studies. Controls did not have follow up serum studies. Furthermore, we did not control for ingestion of vitamins or supplements that might have altered the retinoid and carotenoid values. Finally, interpretation of the findings of this exploratory study needs to account for the multiple statistical tests performed.

Our results further understanding of the roles of vitamin A metabolites and carotenoids in IIH and document lower ATRA in the blood of IIH patients than in the blood of obese controls. The increase in retinoids and pro-vitamin A carotenoids associated with the treatment of IIH with ACZ is suggestive of ACZ having a direct effect on retinoid and pro-vitamin A carotenoid metabolism. Importantly, our data do not support the hypothesis that a “toxic” effect of vitamin A due to increased levels of free retinol or ATRA in blood or CSF is required to develop IIH.

References
